

DeQuanto[®] Adalimumab (Humira[®]) PK ELISA Kit

PK1002

USER MANUAL

Immunoassay for quantitative determination of Adalimumab drug in human serum or plasma

96- Well Microtiter plate

Research Use Only (RUO)

Please read this user's manual carefully before using the kit



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TABLE OF CONTENTS

INTRODUCTION	4
ASSAY PRINCIPLE	4
MATERIALS PROVIDED AND STORAGE.....	4
MATERIALS REQUIRED BY END-USER	5
ASSAY DURATION	5
PRECAUTIONS	5
SPECIMEN COLLECTION AND STORAGE.....	6
SERUM.....	6
PLASMA.....	6
REAGENT PREPARATION	6
WASH BUFFER-A.....	6
WASH BUFFER-B.....	6
ASSAY DILUENT.....	6
ASSAY MATRIX.....	6
TMB SUBSTRATE.....	6
STANDARDS	7
QUALITY CONTROL.....	7
DETECTION ANTIBODY.....	8
ASSAY PROCEDURE.....	8
PRE-PROCESSING OF 96-WELL MICROTITER PLATE.....	8
ANALYTE ADDITION.....	8
DETECTION ANTIBODY.....	8
SUBSTRATE ADDITION AND MEASUREMENT	8
ASSAY SUMMARY	9
CALCULATION OF RESULTS	10
EXAMPLE DATA.....	10
ASSAY CHARACTERISTICS	11
VALIDATION SUMMARY	11
STANDARD CURVE MODEL.....	11

ACCURACY AND PRECISION	11
SELECTIVITY	11
SENSITIVITY	11
DRIFT	11
DILUTION LINEARITY	11
ABBREVIATIONS	12
REFERENCES	13
TROUBLESHOOTING	14
PLATE LAYOUT	15
NOTES	18
CUSTOMIZED SERVICES	21

INTRODUCTION

Adalimumab (Humira®) is a recombinant human IgG1 monoclonal antibody specific for Tumor Necrosis Factor-Alpha (TNF- α) and is used to treat rheumatic arthritis, intestinal disorders, dermatological diseases and cancer. Adalimumab specifically binds to TNF alpha and blocks its interaction with p55 and p75 cell surface TNF receptors and reduces the inflammation and subsequently improves the patient's health.

Drug level quantification can be important to adapt patient prescription or to switch to an alternative TNF inhibitor drug.

EMA Bio-analytical Method Validation Guidelines [2] and industry-recommended practices for ligand-binding assays [3, 4, and 5] were used for validation of this kit. This Adalimumab ELISA kit has been developed for specific quantification of Adalimumab concentration in human serum or plasma with high sensitivity and reproducibility.

ASSAY PRINCIPLE

The DeQuanto® Adalimumab (Humira®) PK ELISA kit is an ELISA based immunoassay. The Adalimumab present in the sample binds to pre coated TNF- α on polystyrene microtiter plate. Non bound or the excess substances are removed by washing. Horseradish Peroxidase (HRP) labeled antibody is added to the wells which binds to the TNF/Adalimumab complex in the wells. Following a wash to remove any unbound antibody-enzyme reagent, substrate solution is added to the wells. A color product is formed in proportion to the amount of Adalimumab present in the sample or standards. The color development is stopped by addition of a stop solution. The absorbance is measured at 450 nm in a microtiter plate reader. The concentration of Adalimumab in a sample can be interpolated from the standard curve.

MATERIALS PROVIDED AND STORAGE

The DeQuanto® Adalimumab (Humira®) PK ELISA kit is recommended to be stored as mentioned in table below.

Sl. No.	Description	Quantity	Volume	Strength/Conc.	Storage
1	Pre Coated 96-well microtiter plate	1 plate	NA	NA	-20°C
2	Adalimumab (Humira®) Standard	1 Vial	10 μ l	50 mg/ml	-20°C
3	Detection Antibody	1 Vial	20 μ l	2000X	-20°C
4	Assay Diluent (Pre-mix powder)	1 Bottle	1 g	NA	2-8°C
5	TMB Substrate	1 Vial	1 ml	20X	2-8°C
6	Wash Buffer-A	1 Bottle	70 ml	20X	2-8°C
7	Plate sealer	2 Nos	NA	NA	RT

MATERIALS REQUIRED BY END-USER

All the following materials are required but not available with the kit.

- Human Serum or Plasma
- Tween-20
- 2N H₂SO₄
- Deionized or ultra pure water
- Microtiter plate reader capable of measuring absorbance at 450 nm (Reference wave length 600 nm is optional)
- Software capable of 4PL or 5PL parameter logistic curve fitting for data analysis

ASSAY DURATION

The approximate time taken for each step of the assay are shown in the table below

Sl. No.	Step	Duration
1	Reagent thawing and preparation	30 minutes
2	Analyte addition and incubation	90 minutes
3	Detection Antibody Addition and Incubation	30 minutes
4	Substrate Addition and Measurement	30 minutes
5	Calculation of Results	15 minutes

PRECAUTIONS

1. User should be trained with ELISA based assays and test procedure
2. All reagents have to be at room temperature before use, except the detection antibody (should be stored at -20°C to ensure the stability)
3. Avoid repeated freeze/thaw cycles for all reagents
4. Handle all reagents wearing gloves and other protective gears
5. Do not pipette any reagents by mouth
6. Use calibrated pipettes and devices only
7. Sodium azide inactivates HRP, do not use sodium azide-containing solutions
8. Any modification in the standard assay procedure may influence the kit performance
9. The described pipetting volume, incubation time, temperature steps should be performed according to the user manual
10. TMB/H₂O₂ Substrate and Detection Antibody are light sensitive, hence should not be exposed to light
11. Avoid physical contact with Stop solution
12. Centrifuge all vials briefly before use
13. Use disposable pipette tips for each transfer to avoid cross contamination
14. Any samples which are out of assay range should be repeated using different dilutions
15. Reagents from different batch/lot are not interchangeable
16. Do not mix reagents from different kit batches/lots
17. The waste disposal should be performed according to your laboratory regulations

SPECIMEN COLLECTION AND STORAGE

SERUM

Use serum clot tube and allow the blood sample to coagulate at room temperature (RT) for 30 minutes. Centrifuge at 5000 RPM for 10 minutes at RT. Aliquot the clear serum and store at -20°C. Avoid repeated freeze/ thaw cycles.

PLASMA

Use K₂ EDTA as anticoagulant for blood collection and allow at RT for 30 minutes. Centrifuge the sample at 5000 RPM for 10 minutes at RT. Aliquot the clear plasma and store at -20°C. Avoid repeated freeze/ thaw cycles.

NOTE: Grossly haemolyzed, lipemic or contaminated sample (serum or Plasma) may lead to inaccurate results and is not recommended to use with this procedure.

It is recommended that the end user must validate assay procedure using either serum or plasma.

REAGENT PREPARATION

WASH BUFFER-A

1. Thaw the wash buffer at RT until it is a clear solution
2. Prepare 1000 ml of 1X wash buffer-A, using 50 ml of 20X Wash buffer-A and make up the volume to 1000 ml with ultra pure water

WASH BUFFER-B

1. Aliquot 500 ml of wash buffer-A
2. Add 0.05% (250 µl) of Tween-20 solution and mix it for 10 minutes on magnetic stirrer at RT

ASSAY DILUENT

1. Add 100 ml of 1X wash buffer-A (W/V) into the assay diluent bottle
2. Allow it to mix on rocker for 10 minutes or by gentle manual mixing at room temperature
3. Assay diluent should appear as a clear solution after mixing
4. Use this assay diluent buffer for assay matrix and test sample dilution

ASSAY MATRIX

1. Dilute human serum or plasma at MRD (1:200) in assay diluent
2. Use this assay matrix for standards and quality control preparation

CAUTION

- Human serum or plasma used for assay matrix must be from healthy volunteers

TMB SUBSTRATE

1. Dilute the 20X TMB substrate to 1X solution in ultra-pure water (600 µl of TMB substrate to 11.4 ml of ultra-pure water)
2. Mix the 1X solution thoroughly on vortex mixer or manually


STANDARDS

1. Prepare a main stock of 1000 µg/ml by diluting the Adalimumab Standard (100 mg/ml) in neat human serum or plasma (5 µl of Adalimumab standard in 495 µl of human serum or plasma)
2. Prepare a sub stock of 5 µg/ml (5000 ng/ml) as given in table below:

Required sub-stock Conc (µg/ml)	Volume of Main-stock to be taken (µl)	Volume of Assay Diluent (µl) to be taken	Final volume (µl)	Dilution factor
5	5	995	1000	1:200

3. Prepare standards 1 to 14 and zero standard (blank) in assay matrix as given in the table below

Sub stock of Adalimumab (ng/ml)	Volume from sub stock (µl)	Assay Matrix Volume (µl)	Final concentration (ng/ml)	Standard No
5000.00	90.00	1410.00	300.00	1
300.00	600.00	300.00	200.00	2
200.00	600.00	300.00	133.33	3
133.33	600.00	300.00	88.89	4
88.89	600.00	300.00	59.26	5
59.26	600.00	300.00	39.51	6
39.51	600.00	300.00	26.34	7
26.34	600.00	300.00	17.56	8
17.56	600.00	300.00	11.71	9
11.71	600.00	300.00	7.80	10
7.80	600.00	300.00	5.20	11
5.20	600.00	300.00	3.47	12
3.47	600.00	300.00	2.31	13
2.31	600.00	300.00	1.54	14
-	-	300.00	0.00	Blank

CAUTION 
<ul style="list-style-type: none"> ▪ <i>Vortex gently to mix during each step</i> ▪ <i>Use appropriate pipette range and do not change pipette in between the assay procedure</i>

QUALITY CONTROL

1. Prepare minimum of three QC samples in assay matrix

DETECTION ANTIBODY

1. Dilute the 2000X Detection Antibody in assay diluent at 1:2000 (take 10 µl of 2000X detection antibody and add to 20 ml of assay diluent)
2. Gently mix the detection antibody before use

ASSAY PROCEDURE

PRE-PROCESSING OF 96-WELL MICROTITER PLATE

1. Thaw the precoated 96-well microtiter plate at room temperature for half an hour
2. Wash the plate with 1X wash buffer-A, allowing 2 minutes for soaking
3. Decant off the contents of the plate

CAUTION

- *Use multichannel pipette*
- *Keep the programs for plate washer ready (if automated washer is used)*

ANALYTE ADDITION

1. Prepare standards, QC samples and test samples
2. Add 100 µl of standards, QC and test or specimen samples to the wells of the microtiter plate as per the plate template
3. Seal plates with adhesive plate sealer
4. Incubate the microtiter plate at RT (~25°C) for 1 hour

CAUTION

- *Refer the sample plate layout given at the end of this user manual, for standards and control*
- *Before addition of samples to the microtiter plate, keep the Plate template ready.*

DETECTION ANTIBODY

1. Discard the contents of each well and wash 3 times with 1X wash buffer-B and followed by 3 washes with 1X wash buffer-A, allowing 2 minutes for soaking between each wash step
2. Blot the microtiter plate on absorbent paper to remove any residual reagent from the wells
3. Add 100 µl of Detection Antibody solution to each well of the microtiter plate
4. Seal the microtiter plate with adhesive plate sealer
5. Incubate the microtiter plate at RT (~25 °C) for 30 minutes

SUBSTRATE ADDITION AND MEASUREMENT

1. Discard the contents of each well and wash 3 times with 1X wash buffer-B and followed by 3 washes with 1X wash buffer-A, allowing 2 minutes for soaking between each wash step
2. Blot the microtiter plate on absorbent paper to remove any residual reagent from the wells
3. Add 100 µl/well of 1X TMB substrate solution and incubate the plate in dark at RT (~25°C) for 15 minutes
4. Add 50µl/well of Stop solution to stop the reaction
5. Read the absorbance in micro plate reader set to 450 nm, set the reference wavelength to 600 nm

CAUTION

- *Add stop solution in the same order of addition of TMB substrate to the plate*

ASSAY SUMMARY

1. Prepare all reagents as instructed in the user manual



2. Add 100 µl of standard, quality control and test/specimen samples to each well of microtiter plate



3. Seal the microtiter plate with adhesive plate sealer and incubate at RT (~25°C) for 1 hour



4. Decant the contents of the microtiter plate and wash the microtiter plate 3 times with wash buffer-B, followed by 3 times with wash Buffer-A, allowing 2 minutes of soaking between each wash step



5. Blot the plate on absorbent paper to remove any residual reagent from the wells



6. Add 100 µl of detection antibody to each well of the microtiter plate and incubate for 30 minutes at RT (~25°C)



7. Repeat the steps 4 and 5



8. Add 100 µl of 1X TMB solution to each well of the microtiter plate and incubate for 15 minutes in dark at RT (~25°C)



9. Add 50 µl of stop solution (2N H₂SO₄) to each well



10. Read the absorbance at 450 nm and 600 nm as reference wave length

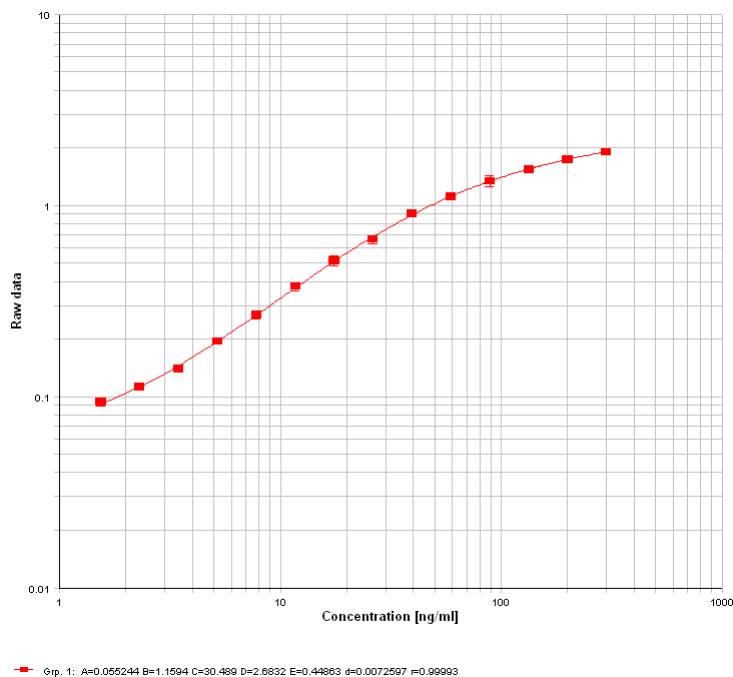
CALCULATION OF RESULTS

After the absorbance is read at 450 nm and 600 nm as reference wave length, construct a standard curve of difference data using software capable of generating four or five parameter logistic (4PL or 5PL) curve fit. Absorbance of the test/specimen and the QC samples are interpolated from the standard curve. Report the values of test/specimen samples within the assay range.

EXAMPLE DATA

This standard curve results are provided for demonstration purpose only.

The standard curve should be run for each assay.



Standard	Conc (ng/ml)	Mean Abs
S1	300.00	1.907
S2	200.00	1.743
S3	133.33	1.548
S4	88.89	1.338
S5	59.26	1.114
S6	39.51	0.908
S7	26.34	0.666
S8	17.56	0.513
S9	11.71	0.377
S10	7.80	0.268
S11	5.20	0.194
S12	3.47	0.140
S13	2.31	0.112
S14	1.54	0.094
Blank	0.00	0.046

ASSAY CHARACTERISTICS

VALIDATION SUMMARY

Denovo Biolabs have validated this assay in human serum matrix according to EMA Bio-analytical Method Validation Guidelines [2] and industry-recommended practices for ligand-binding assays [3, 4, and 5]. However, such validation is generic in nature and it is intended to only supplement but not substitute specific validation as required by regulations or otherwise in each case.

STANDARD CURVE MODEL

The calibration standards were generated by spiking Adalimumab in human serum at MRD 1:200. The standard curve consisted of twelve non-zero standards (from 300 ng/ml to 1.54 ng/ml). A 5PL model was used to fit the standard curve. The regression model was accepted as the %RE of the back-calculated value for at least 75% calibrators was within 20% of nominal concentration, except at the ULOQ and LLOQ where it was within 25%.

ACCURACY AND PRECISION

Accuracy (%RE) of all QC samples was within 20% across all batches. Inter-assay and pooled (cumulative) intra-assay precision (%CV) of each QC sample was $\leq 20\%$. The total error was within 20%.

SELECTIVITY

Ten different human plasma matrices were tested for selectivity in a single batch experiment by recovery studies at two QC points. 80% of 10 different individual plasma samples passed the acceptance criteria of %RE within 20% except at LLOQ where it was within 25%.

SENSITIVITY

In all batch runs LLOQ-QC at 5.2 ng/ml showed %RE within the 20% range confirming the sensitivity of the assay to be 5.2 ng/ml.

DRIFT

The drift parameter was evaluated by placing QC samples at different positions of the 96-well microtiter plate in an intra-assay batch. All the three QCs samples passed the drift criteria and the %CV of all the QC samples were within $\leq 20\%$, except LLOQ-QC where it was within $\leq 25\%$.

DILUTION LINEARITY

From the sub-stock a 1000X, ULOQ sample was made which was further diluted into other QC samples. Each of these QC samples was tested for their %RE values. All near QC samples passed the acceptance criteria. 1000X-ULOQ did not pass the criteria due to Hooks Effect.

ABBREVIATIONS

RT	Room Temperature
Abs	Absorbance
Conc.	Concentration
HRP	Horse Radish Peroxidase
PK	Pharmaco Kinetics
TMB	3, 3', 5, 5'-Tetramethylbiphenyl-4, 4'-Diamine
TNF	Tumor Necrosis Factor
ELISA	Enzyme Linked Immunosorbent Assay
%CV	Coefficient of variations
%RE	Relative Error
MRD	Minimum Required Dilution
PL	Parameter Logistic
QC	Quality Control
CTRL	Control
ULOQ	Upper Limit of Quantification
HQC	High Quality Control
LQC	Lower Quality Control
LLOQ	Lower Limit of Quantification
°C	Degree Celsius
µg	Microgram
mg	Milligram
µl	Microliter
Eg.	Example
ml	Milliliter
W/V	Weight by Volume
IgG	Immunoglobulin
ng	Nanogram
nm	Nanometer
2N H ₂ SO ₄	2 Normal Sulphuric Acid
K ₂ EDTA	Ethylene Di-amine Tetra Acetic acid (Di-Potassium salt)
RPM	Revolutions Per Minute

REFERENCES

1. *Guideline on bioanalytical method validation*; 21 July 2011
EMA/CHMP/EWP/192217/2009 Committee for Medicinal Products for Human Use
(CHMP)
2. DeSilva *et al* (2003), *Recommendations for the Bioanalytical Method Validation of Ligand-binding Assays to Support Pharmacokinetic Assessments of Macromolecules*, *Pharmaceutical Research*, **20 (11)**: 1885-1900.
3. Findlay *et al.* (2000) *Validation of Immunoassays for bioanalysis: A pharmaceutical industry perspective*. *J. Pharm. Biomed. Anal.* **21**, 1249-73.
4. Kelley and DeSilva (2007); *Key Elements of Bioanalytical Method Validation for Macromolecules*, *AAPS Journal*, **9 (2)**, E156-E163.
5. European Medicines Agency, *EPAR – Product Information on MabThera*, Last updated 12 June 2012, available at www.ema.europa.eu.

TROUBLESHOOTING

Problem	Probable Causes	Solution
High or Low OD value of test samples	Test samples contain analyte concentrations greater than assay range	Repeat the assay with multiple dilutions of the test samples
	Test Samples contain no or below detectable levels of analyte	Test sample contains the drug level below the LLOQ
High % CV (High variability in duplicate OD values)	Unequal volumes	Ensure the calibrated pipettes Ensure pipette tips are tightly secured while adding the solution to each well
	Inadequate washing	Ensure the wash system is working properly if the automated wash station is used. Also ensure the manual washes
	Non-homogenous solution	Ensure that all solution prepared are adequately mixed.
	Edge effect	Use plate sealer and maintain the temperature as recommended
	Cross-well contamination	Ensure the good pipetting practice
	Inadequate mixing of TMB substrate solution	Ensure the TMB substrate solution is mixed thoroughly before use
Low signal	Standard was incompletely reconstituted or the storage was inappropriate	Reconstitute standard according to the user's manual Ensure the kit reagents are not expired
	Reagents added to wells with incorrect concentrations	Cross check for calculation and pipetting errors
	Incorrect incubation time or temperature	Adhere to the recommended assay procedure
Standard curve with R ² value <0.95	Pipetting error	Ensure the pipetting and the analyst training

PLATE LAYOUT

	12												
	11												
	10												
	9												
	8												
	7												
	6												
	5												
	4	S1	S9	S2	S10	S3	S11	S4	S12	S5	S13	S6	S14
	3	S1	S9	S2	S10	S3	S11	S4	S12	S5	S13	S6	S14
	2	S1	S9	S2	S10	S3	S11	S4	S12	S5	S13	S6	S14
	1	S1	S9	S2	S10	S3	S11	S4	S12	S5	S13	S6	S14
A		S1	S9	S2	S10	S3	S11	S4	S12	S5	S13	S6	S14
B		S2	S10	S3	S11	S4	S12	S5	S13	S6	S14	Ctrl	
C		S3	S11	S4	S12	S5	S13	S6	S14	Ctrl			
D		S4	S12	S5	S13	S6	S14	Ctrl					
E		S5	S13	S6	S14	Ctrl							
F		S6	S14	Ctrl									
G		S7	Ctrl										
H		S8											

	1	2	3	4	5	6	7	8	9	10	11	12
A	S1	S1	S9	S9								
B	S2	S2	S10	S10								
C	S3	S3	S11	S11								
D	S4	S4	S12	S12								
E	S5	S5	S13	S13								
F	S6	S6	S14	S14								
G	S7	S7	Ctrl	Ctrl								
H	S8	S8										

NOTES

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CUSTOMIZED SERVICES

PRECLINICAL/CLINICAL SAMPLE ANALYSIS

ELISA based validation and clinical or pre-clinical sample analysis.

POLYCLONAL ANTIBODY DEVELOPMENT

PAb development against peptide or protein, peptide designing, high titer antibody purification, characterization.

MONOCLONAL ANTIBODY DEVELOPMENT

MAB development against peptide or protein, peptide designing, best reacting/stable hybridoma development, sub cloning, small scale/large scale hybridoma culture and antibody purification, characterization.

CLONING, PROTEIN EXPRESSION PURIFICATION

Gene synthesis, site directed mutagenesis, cloning and expression in bacterial systems and other customized molecular biology services.

ANTIBODY CONJUGATION

With HRP, FITC, Biotin, ALP and many more molecules.

CELL BASED ASSAYS

Cell based assays, neutralization assays and potency ratio estimation of drugs with reference drugs, drug efficacy assessment and customized in-vitro, in-vivo assay development.

CUSTOMIZED ELISA BASED ASSAY/METHOD DEVELOPMENT

Different formats of ELISA based assay development.

For any queries/enquiries related to our products or services, please contact us by mailing us at info@denovobiolabs.com or call us at +91-80-29575711